

Dominican Scholar

Natural Sciences and Mathematics | Faculty Scholarship

Department of Natural Sciences and Mathematics

1-1-2017

Steroid hormone receptors as prognostic markers in breast cancer.

Maggie Louie

Department of Natural Sciences and Mathematics, Dominican University of California, maggie.louie@dominican.edu

Mary B. Sevigny

Department of Natural Sciences and Mathematics, Dominican University of California, mary.sevigny@dominican.edu

Survey: Let us know how this paper benefits you.

Recommended Citation

Louie, Maggie and Sevigny, Mary B., "Steroid hormone receptors as prognostic markers in breast cancer." (2017). *Natural Sciences and Mathematics | Faculty Scholarship*. 40. https://scholar.dominican.edu/natural-sciences-and-mathematics-faculty-scholarship/40

This Article is brought to you for free and open access by the Department of Natural Sciences and Mathematics at Dominican Scholar. It has been accepted for inclusion in Natural Sciences and Mathematics | Faculty Scholarship by an authorized administrator of Dominican Scholar. For more information, please contact michael.pujals@dominican.edu.

Review Article Steroid hormone receptors as prognostic markers in breast cancer

Maggie C Louie*, Mary B Sevigny*

Department of Natural Sciences and Mathematics, Dominican University of California, 50 Acacia Avenue, San Rafael, CA 94901, USA. *Equal contributors.

Received June 26, 2017; Accepted July 5, 2017; Epub August 1, 2017; Published August 15, 2017

Abstract: Despite the existence of many promising anti-cancer therapies, not all breast cancers are equally treatable, due partly to the fact that focus has been primarily on a few select breast cancer biomarkers- notably ER α , PR and HER2. In cases like triple negative breast cancer (ER α , PR, and HER2·), there is a complete lack of available biomarkers for prognosis and therapeutic purposes. The goal of this review is to determine if other steroid receptors, like ER β and AR, could play a prognostic and/or therapeutic role. Data from various *in vitro*, *in vivo*, and clinical breast cancer studies were examined to analyze the presence and function of ER β , PR, and AR in the presence and absence of ER α . Additionally, we focused on studies that examined how expression of the various steroid receptor isoforms affects breast cancer progression. Our findings suggest that while we have a solid understanding of how these receptors work individually, how they interact and behave in the presence and absence of other receptors requires further research. Furthermore, there is an incomplete understanding of how the various steroid receptor isoforms interact and impact receptor function and breast cancer progression, partly due to the difficulty in detecting all the various isoforms. More large-scale clinical studies must be made to analyze systematically the expression of steroid hormone receptors and their respective isoforms in breast cancer patients in order to determine how these receptors interact with each other and in turn affect cancer progression.

Keywords: Breast cancer, steroid hormone receptors, prognostic markers, estrogen, progesterone, androgen

Introduction

Though both estrogen and progesterone receptors are commonly used as prognostic markers for breast cancer, current endocrine therapy primarily targets the estrogen receptor, ERa. Unfortunately, for about 10-15% of breast cancer patients [1-3]- like those diagnosed with triple negative breast cancer, defined as breast tumors lacking the expression of estrogen receptor alpha (ERα), progesterone receptor (PR) and human epidermal growth factor receptor type 2 (HER2), ERα-/PR-/HER2- - the established endocrine therapies are ineffective, highlighting an urgent need for additional therapeutic targets in breast cancer. Therefore, the goal of the following review is to examine the role of the steroid hormone receptors- ERα, ERβ, PR, and androgen receptor (AR)- in the progression of breast cancer in order to determine their role and utility as prognostic markers and therapeutic targets.

Steroid receptors: an overview

The steroid hormone receptor subfamily, which includes estrogen receptor (ER), progesterone receptor (PR), androgen receptor (AR), and glucocorticoid receptor (GR), is part of the larger superfamily of nuclear hormone receptors [4]. The members of this superfamily function as ligand-gated transcription factors that modulate the expression of genes [5]. While unbound steroid receptors are typically located in the cytosol, ligand binding induces receptor dimerization and conformational changes which in turn exposes the nuclear localization signal, allowing translocation into the nucleus [6]. Once inside the nucleus, the receptor dimer recognizes and binds specific DNA sequences

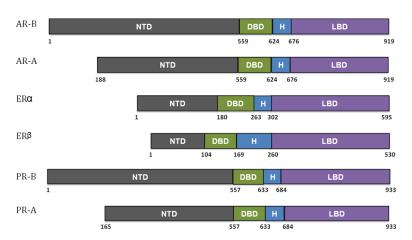


Figure 1. Comparing the functional domains of nuclear hormone receptors. *NTD*, N-terminal A/B hypervariable domain; *DBD*, DNA binding domain; *H*, hinge region; *LBD*, ligand binding domain. Adapted from [196].

that in turn results in enhancing or silencing the transcription of specific target genes regulated by the receptor [7, 8].

As depicted in Figure 1, nuclear hormone receptors share common functional domains, such as a DNA binding domain (DBD), a ligand binding domain (LBD), and two transactivation domains (AF-1 and AF-2) [9-12]. DBDs contain two zinc finger motifs that allow them to recognize and bind to specific DNA sequences- often referred to as hormone response elements (HREs)- within the promoter and/or enhancer regions to regulate transcription. Different steroid hormone receptors bind to different response elements, thus allowing the receptors to regulate subsets of genes that are necessary to elicit a physiological response. The LBD is involved in the binding of specific hormones, which induces dimerization and nuclear translocation [9-12]. The two transactivation domains, AF-1 and AF-2, are important for modulating transcription of the target genes. AF-2 is located within the LBD and is involved in liganddependent transactivation, while AF-1 is found in the N-terminal A/B hypervariable domain (NTD) and is responsible for ligand-independent transcriptional activation and mediates protein-protein interaction with other transcription factors [13]. AF-1 is also responsive to phosphorylation by kinases that are activated in various signaling pathways, including the epidermal growth factor receptor (EGFR) pathway.

Estrogen receptor

The estrogen receptor (ER) was identified in the 1950s by Dr. Elwood V. Jensen as reviewed in [14, 15]. Eventually, it was determined that three forms of ER exist- $ER\alpha$, ERβ, and GPR-30 [16-19]. All three ERs are encoded by different genes located on different chromosomes [20]. ERα is encoded by the gene ESR1 located on chromosome 6; ERB is encoded by gene ESR2 on chromosome 14; and GPR-30 is encoded by the GPER gene on chromosome 7. While ERa and

ERβ are nuclear hormone receptors, GPR-30 is not a member of the steroid receptor family but is instead a G-protein coupled receptor that has been shown to bind and respond to estrogen [21-23]. Therefore, since our interest is in the role of steroid hormone receptors in breast cancer, GPR-30 will not be discussed further in this review.

Although ERα and ERβ are both expressed in breast tissue and bind to estrogen with similar affinities [24], studies have shown that only ERα is necessary for normal mammary gland development [25, 26] leading researchers to question the function of ERB in normal breast tissue. Multiple studies have also reported that ERβ expression actually represses ERα expression and function [27, 28]. In addition to the breast, both nuclear receptors are expressed in many other tissues within the human body, including the endometrium, ovary, testes, cerebral cortex, myocardium, and thyroid [29-32]. However, their expression patterns do differ in certain tissues- for example, ERa is the sole estrogen receptor expressed in the hippocampus, and only ERB is found in prostate tissue [32].

In addition to full length ER α (as shown in **Figure 1**), there are 2 truncated splice variants of ER α - 46 kDa estrogen receptor (ER46) [33] and 36 kDa estrogen receptor (ER36) [34, 35]. Some of the full length ER α (ER66) along with ER36 and ER46 [36] associate with the plasma

membrane and are referred to as mERs due to their ability to translocate to the plasma membrane through palmitoylation and caveolin-1 association, which in turn allows for rapid estrogen receptor signaling. ER46 does not have an AF-1 transactivation domain, which suggests that ER46 does not mediate any nuclear functions. ER36 lacks both AF-1 and AF-2 transactivation domains, and part of the ligand binding domain is replaced by a 27 amino acid sequence in the C terminus [34]. ER36 has been shown to mediate estrogen stimulation via mitogen-activated protein kinase (MAPK) pathway [35].

As with ER α , ER β also exists as several isoforms- ER β 1 (**Figure 1**), ER β 1 "short form", ER β 2/cx, ER β 3, ER β 4, and ER β 5 [37-40]. Most are due to the alternative splicing of exons 7 and 8, although the truncated form of ER β 1 is due to proteolysis at the N-terminus [41, 42]. Of all the isoforms, only ER β 1 contains the ligand-binding domain [43]. However, it has been shown that the formation of heterodimers between ER β 1 and the other isoforms increases the transcriptional activity of ER β 1 [37].

Progesterone receptor

The expression of PR is primarily regulated by ERα at the transcriptional level [44, 45]. There are two known isoforms of PR, PR-A and PR-B. PR-A is a truncated version of PR-B, lacking 164 amino acids at the N-terminus (Figure 1). The two proteins are transcribed from two different promoters located within the same gene on chromosome 11 [46] and can form homo-or heterodimers. Studies using knockout mice confirmed the functional importance of both PR isoforms [47, 48]. Although animals lacking PR-A did not display significant developmental effects in the mammary glands or thymus, they did display severe dysfunctions in their ovaries and uterus resulting in infertility, suggesting that its primary function is maintaining normal ovarian and uterine functions [49-52]. Converselv. PR-B knockout-mice retained normal ovarian, uterine, and thymic functions but exhibited a significant decrease in mammary ductal morphogenesis [49-51, 53], indicating that PR-B mediates the proliferative effects of progesterone in the mammary gland.

Structural and functional analyses of each PR isoform suggest that they have different tran-

scription activation properties when bound to progesterone [53, 54]. According to Richer et al., approximately 27% of PR-regulated genes are controlled by both PR isoforms. However, this study also indicated that PR-B alone controls the majority of the PR-regulated genes in comparison to PR-A alone (69% versus 4%) [55] which may be in part due to PR-B being intrinsically a stronger transcriptional activator than PR-A. In fact, PR-A has been reported to function as a transcriptional repressor under certain cellular conditions [56]. Curiously however, it is PR-A- not PR-B- that is more frequently over-expressed in breast cancer [57]. Some studies have even indicated that it is not so much the expression of either isoform but rather the ratio of the two isoforms that are important in breast cancer development. For example, a higher ratio of PR-A/B has been associated with poorer prognosis and response to hormone therapy [58].

Androgen receptor

AR is expressed in all tissues, including testis, prostate, foreskin, cervix, vagina, mammary glands, bone, brain, sebaceous and sweat glands of the skin, and breast [59-61]. The gene that codes for the androgen receptor (AR) is located on the X chromosome [60, 62]. While AR is closely related to PR and ER, one distinctive feature of the AR protein is the presence of glutamine and glycine repeats in the N-terminal activation domain of the receptor which have been linked to certain cancers and chronic neurological diseases in humans [60, 63, 64]. In 1994, Wilson and McPhaul discovered two isoforms of AR in human genital skin fibroblasts that are structurally very similar to PR-A and PR-B [65]. Their formation is due to two distinct translation initiation sites which result in the full-length receptor (110 kDa) and an N-terminally truncated form (87 kDa) known as AR-B and AR-A, respectively (Figure 1). Since 1994, several low molecular weight isoforms of AR have been identified, particularly in prostate cancer cell lines and tumors (reviewed in [66]). A few different mechanisms are responsible for these variants, including premature chain termination during translation, proteolysis by calpains, and alternative splicing [66]. Several isoforms, such as AR-V7 (also known as AR3), lack the LBD and are thus capable of transcription activation in the absence of androgen (i.e.

androgen-independent transcription) [67, 68]. Though originally found in prostate cancer cases that have become androgen deprivation therapy (ADT) or castration resistant [67-70], AR-V7 has likewise been found in a substantial number of primary breast tumors (~50%) and most breast cancer cell lines [71]. Another AR variant, Δ3AR, has been found exclusively in some breast tumors and breast cancer cell lines but not in normal breast tissue [72]. Originally discovered in patients suffering from androgen insensitivity syndrome (AIS), the Δ3AR isoform lacks the second zinc finger in the DBD, which potentially could reduce its ability to inhibit cell growth, at least within an ERα+ setting [72, 73].

ERα-positive breast cancer

Prognosis and treatment

In 1896, George Thomas Beatson found that removing the ovaries from patients with advanced stages of breast cancer resulted in significant regression, which later lead to the speculation of estrogen's stimulating effect on breast cancer. Hence, oophorectomy and/or the use of drugs that target the estrogen receptor have become standard therapies for treating estrogen responsive breast cancer. To date, much of what we know about the relationship between ER and breast cancer centers primarily on one particular receptor- ERα. In fact, it is the presence or absence of ERa that determines whether a patient's breast cancer can be classified as either estrogen receptor positive or negative, respectively. ERα-negative breast cancers may express other hormone receptors such as PR, AR, and even ERβ, but they are often non-responsive to estrogen. Of all the breast cancer subtypes, ERα-positive breast cancer is the most prevalent, accounting for approximately 75% of breast cancers diagnosed in women [1].

Studies have shown that patients with ER α -positive breast cancers have a better prognosis because these tumors tend to be lower grade and have less aggressive phenotypes. Even patients with metastatic tumors that expressed ER α often had significantly better survival outcomes in comparison to patients with ER α -negative tumors [74], and this is most likely due to the fact that most patients with ER α -positive

tumors also had an increase likelihood of responding to the established endocrine therapies [74]. However, not all ER-positive tumors respond to endocrine therapy, and even those that are initially responsive eventually become resistant as the disease progresses.

Drugs that specifically target ERα- estrogen receptor antagonists- can be used to treat or manage the disease. ER antagonists specifically compete with estrogen and block it from binding to the receptor. There are two types of ER antagonists- 1) selective ER modulators (SERMs), also referred to as partial antagonists, and 2) pure or complete antagonists, also referred to as selective ER down-regulators (SERDs) [75]. Among the most common SERMs are the anti-estrogens tamoxifen and raloxifene. Tamoxifen is effective in antagonizing estrogen-dependent cancer cell growth by binding to ERa and promoting the recruitment of co-repressors rather than co-activators in mediating transcriptional repression of ER target genes [11, 76]. An example of a pure antagonist or SERD is ICI 182, 780, also referred to as faslodex or fulvestrant, which binds to either ER α or β and promotes receptor degradation [77-79]. Ultimately, ERα is considered a good prognostic marker for breast cancer not only because it is vital in both the development and progression of the disease but also because its presence determines whether the cancer will likely respond to anti-estrogen treatment.

Association with progesterone receptors

Of all the ERα+ mammary tumors, about 50-60% are PR+ [80-82]. Yet while the ER status has been well established as a predictive factor for breast cancer prognosis and cancer treatment, less is known about the significance of PR in the presence of ERa. Studies have shown that tumors expressing both receptors tend to be less aggressive and least likely to metastasize [83, 84]. Others have confirmed that the presence of both ER α and PR in tumors often translates to better prognosis [83-86]. Specifically, Dunnwald et al. examined the correlation between PR/ERα expression and mortality risk amongst 155,175 breast cancer patients and found that patients with ERα+/PR+ tumors had lower mortality rates compared to women with $ER\alpha^+/PR^-$, $ER\alpha^-/PR^+$, and $ER\alpha^-/PR^-$. The highest mortality rate was seen in ERα⁻/PR⁻

patients. A similar study carried out by Salmen et al. determined that patients with ER α /PR tumors had worse prognoses than patients with ER α /PR+ or ER α +/PR+ tumors [87]. Furthermore, tumors that have lost PR expression are often more aggressive and have negative prognoses, signifying that PR is an important indicator of the progression of the disease [82].

Compared to ERα⁺/PR⁺ patients, a smaller percentage of ERα⁺/PR⁻ breast cancer patients respond to tamoxifen treatment [85, 88-91], suggesting that PR plays an important role in endocrine therapy response. This could account for the consistent observation that not all ERα⁺ breast cancer patients respond to endocrine therapies like tamoxifen. As noted earlier, the presence of ER α may not always be sufficient to indicate positive outcome towards endocrine based-therapeutics. This may be due in part to the fact that expression of the ERa protein does not always translate to a functional ERa signaling pathway. Since PR is one of the target genes of ERa, it has long been proposed that the expression of PR may serve as a good indicator of ERa functionality and signaling [92]. Studies have in fact confirmed that the presence PR in ERα+ breast cancer significantly improves the outcome prediction for adjuvant endocrine therapy [85, 91, 93-96]. Specifically, Ferno et al. found that patients with ERα⁺/PR⁺ tumors had significant increase in response to adjuvant tamoxifen therapy compared with patients with ERα⁺/PR⁻ tumors [94, 97]. Other studies have also confirmed that PR status can be a better indicator of tamoxifen response than ERa status alone [96, 98]. We speculate that the presence of ERa without PR expression likely suggests a signaling dysfunction in the ERa pathway that reduces its ability to transcriptionally regulate its target genes; therefore, it is not surprising that the presence of both ERa and PR is a better indicator of endocrine therapy responsiveness.

Studies have also demonstrated that PR is associated with overall survival of cancer patients [85, 99]. Specifically, patients diagnosed as $ER\alpha^+/PR^+$ had less cancer recurrence in comparison to $ER\alpha^+/PR^-$ cancer patients [85]. We acknowledge that this reported decrease in breast cancer recurrence in $ER\alpha^+/PR^+$ patients contradicted earlier studies [86,

100], but this may be attributed to the researchers' use of biochemical assays to measure PR, which lack the sensitivity of other methods such as immunohistochemistry, which is the current practice.

Further complicating the analysis of PR's role in breast cancer prognosis and therapeutic response is the existence of the two PR isoforms mentioned previously- PR-A and PR-B. The ratio of PR-A to PR-B has been shown to change during the development of breast cancer [55, 101], and this may alter prognosis and therapeutic response. Although some breast cancers may express both isoforms, the ratios of the two receptors vary, with PR-A showing a higher expression in most tumors [55, 101]. This ratio of PR-A to B can impact the prognosis and staging of the disease, with an equal to low PR-A:PR-B ratio associated with lower tumor grading (G1 and G2) and a high PR-A:PR-B ratio associated with undifferentiated, higher grade tumors (G3) [101]. Recent microarray analysis also suggests that the PR-A:PR-B ratio is a critical determinant of PR target gene selectivity and response to hormonal stimuli [102]. This indicates the importance of evaluating the interplay between PR-A and PR-B when determining the clinical outcomes and responsiveness to endocrine therapy. However, typical methods used to determine the presence of PR in tumors are either ligand binding assays using tumor extracts or antibody-based assays such as immunohistochemistry [103-106], and neither of these assays is capable of differentiating between the two PR isoforms. In some cases, immunohistochemistry may only detect PR-A and not PR-B in formalin fixed tissue, suggesting that conformational differences between the two may interfere with detection [105]. Ultimately, further analysis of the role of PR-A and PR-B in breast cancer is needed to provide a better understanding of PR in prognosis and therapeutic response.

Association with ERB

ER β is not commonly used as a prognostic marker for breast cancer, partially because the presence and function of ER β in ER α -positive tumors are not well understood. As with PR, the existence of several isoforms of ER β adds to the confusion and has resulted in conflicting data, since the majority of studies that analyze

the role of ER β in ER α ⁺ cells do not differentiate between the various ERB isoforms. Despite this complexity, some researchers have had the foresight to study the specific ERB isoforms, and several agree that expression of the ligandbinding isoform ERβ1 in ERα+ cells counteracts ERα activity, thereby suppressing cell proliferation and enhancing apoptosis [39, 107, 108]. Two other studies carried out by Ogawa et al. and Peng et al. found that overexpression of the isoform ER\$2/cx inhibited the transcriptional activity of ERα [38, 109], while this same isoform has been shown to enhance the transcriptional activity of ERB1 [37]. An in vitro study led by John Hawse concluded that the tamoxifen metabolite endoxifen worked best at inhibiting estrogen-mediated cell proliferation in ERα+ cells if ERB was also expressed [110]. It was also determined that endoxifen treatment of cells led to ERB accumulation and subsequent increase in ERα/ERβ heterodimers, providing further evidence that the observed cancer suppressing activity of ERB is due in part to a direct interaction between ERB and ERa. However, a clinical study analyzing the effect of a 2-year tamoxifen treatment on 353 patients with stage II primary breast tumors found that patients who were ERα⁻/ERβ⁺ had significantly greater distant disease-free survival compared with patients who were ERα+/ERβ+, suggesting a mechanism for ERB independent of ERa [111]. Since neither the levels of the tamoxifen metabolite endoxifen nor the exact ERB isoforms were determined in these patients, it is difficult to directly compare this study with that of Hawse's. Yet these seemingly differing conclusions do point to a rather complex role for ERB in breast cancer, which clearly involves more than just inactivation of ERa.

Association with androgen receptors

AR expression is most commonly associated with prostate cancer, and prostate tumor progression is as dependent on AR activity as breast tumor progression is on ER α activity. In fact, the treatment of prostate cancers usually involves hindering AR function via ligand depletion, treatment with AR antagonists, or both [112]. The role of AR in breast cancer is not quite as clear. AR is frequently expressed in ductal carcinoma in situ (DCIS) and invasive breast carcinoma [113]. In addition, most ER α + breast cancers also appear to express AR, as

exemplified by a study carried out by Hu et al. which found that 88% of 1,164 ER positive breast cancer cases also expressed AR [114]. Another study by Agrawal et al. analyzed the importance of using AR as a prognostic marker in 488 breast cancer patients who underwent radical mastectomies [115]. Data from this study suggest that the presence of AR increased the success of adjuvant therapy and prognosis in patients. Additionally, they found that 50.7% of breast cancer patients who were AR negative had lower 5-year survival rates, indicating poorer prognosis [115]. In yet another study carried out by Qu and colleagues, 109 breast cancer patients in Shanghai were retrospectively analyzed between 2003 and 2008 [116]. Of the 109 patients, 52 were diagnosed AR+. Overall, there were 13 deaths and 15 recurrences but only two of the deaths and three of the recurrences were from the AR+ group, which again indicates that AR could be a good marker for longer overall survival and lower risk of recurrence. Conversely, mammary tumors that do not express AR have been shown to respond poorly to hormone therapy [117]. The absence of AR has also been correlated with higher levels of Ki-67, a cell proliferation marker associated with cancer progression [118], though the molecular mechanism behind this finding has not been elucidated. Furthermore, androgen-activated AR appears to directly bind to the ERB promoter and enhance transcription of ERβ in both ERα-positive and -negative breast cancer cells [119], suggesting that blocking AR function may subsequently decrease ERβdependent gene expression.

Although many studies appear to support the hypothesis that AR helps counteract the tumorigenic effects of ERa, others- such as one carried out by Paliouras and Diamandis- report a synergistic mechanism between the two receptors, resulting in an increase in breast cancer progression [120]. Specifically, the ERdependent expression of a group of cancer biomarkers called kallikrein (KLKs) genes was shown to be enhanced significantly by the binding of androgen to AR. One noteworthy limitation of this study is that it focused on just one breast cancer cell line (BT474) and therefore may not be indicative of most breast cancer cases. Another study by Liao et al. showed that simultaneous treatment with androgen and estrogen stimulated mammary gland carcinomas in 100% of the Noble rats tested [121]. The researchers concluded that high levels of androgen and estrogen together may be an important risk factor for breast cancer and that direct binding of androgens to either AR or PR were involved in this carcinogenic process. However, although the researchers analyzed various isoforms of ER and PR, they only concentrated on the two AR isoforms, AR-A and AR-B, and used only male rats in their study. Additionally-and perhaps most importantly- the authors did not sufficiently rule out the possibility that the results were due to aromatization of androgen to estrogen. A more recent study carried out by Richer and colleagues did determine that treatment of AR⁺/ERα⁺ breast cancer cell lines with the AR inhibitor enzalutamide- a drug currently used to treat metastatic prostate cancer- effectively inhibited cell proliferation both in vitro and in animal models, suggesting that AR activity may indeed promote cancer progression in the presence of ERa [122]. Yet again, isoforms of AR were neither analyzed nor even acknowledged in this study.

In contrast to the above studies, several clinical studies have indicated that administration of normal, physiological levels of androgen to women receiving estrogen therapy actually decreases breast cancer risk (reviewed in [59]), but the mechanism behind this- particularly in regards to AR signaling- is not clear. Agrawal and colleagues reported that only in the absence of estrogen will androgens directly bind to ERα and stimulate the proliferation of cancerous cells [115]. Finally, in an excellent, in-depth review of AR in breast cancer, K. M. McNamara and colleagues acknowledge that the role of AR in breast cancer risk and progression depends greatly on the specific disease subtype and the presence or absence of the other steroid receptors, such as ERa [123]. The authors further assert that in the presence of ERα, AR activation appears to counteract disease progression in most breast cancer subtypes. However, a further complication involves the ratio of AR to ER, which also appears to affect progression and efficacy of endocrine treatment. A clinical analysis of 192 ERα+ breast cancer patients carried out by Richer and colleagues revealed that a high AR to ERa ratio correlated positively with increased incidence of tamoxifen failure [122]. Despite all the confusing and contradictory findings, it does appear evident that AR could serve as an important prognostic indicator as long as AR isoforms, ER α status and estrogen levels are also taken into account.

ERα-negative breast cancer

ERβ-positive breast cancer

Although studies have found that both mRNA and protein levels of ERB are significantly lower in breast tumors compared to normal breast tissue [124, 125], approximately 17% of primary breast cancers are ERα-negative/ERβpositive [126, 127]. In addition, 47-60% of all ERα-negative tumors have been reported to be ERβ-positive [42, 126]. Most studies agree that the presence of ERB in these tumors is correlated with a positive prognosis, as the absence of ER β in ER α -negative patients (ER α -/ER β -) is associated with early relapse [111, 128-130]. It has also been reported that as breast tumors become more malignant, ERB expression decreases (reviewed in [131]). However, one recent study carried out by Chen et al. contradicts these findings [132]. The researchers found that expression of ERB actually enhanced cancer progression by inducing the expression of IL-8, which is known to play a role in angiogenesis and metastasis [132, 133].

Predictably, most of the aforementioned studies failed to differentiate between the various isoforms of ERB, which once again could be responsible for these conflicting results. The small handful of studies that actually have analyzed the expression of specific ERB isoforms have found that ER\$1, -2/cx, -3, and -5 are expressed at varying degrees in breast tumors and breast tumor cell lines [42, 43, 134, 135]. ERβ2/cx (also designated simply as ERβ2) is the main isoform expressed in the hormonesensitive breast tumor cell line T47D, whereas ERB5 is the major isoform in the hormoneinsensitive BT20 breast tumor cell line [134]. Increased expression of both ER\$2/cx and ERB5 relative to the full-length isoform ERB1 appears to correlate with increased breast cancer progression [135], and yet increased expression of ERB5 has also been positively correlated with breast cancer survival [136, 137]. A recent clinical study of 95 patients with ERα-negative invasive breast carcinomas demonstrated a correlation between prognosis and the ER\$1 to ER\$2 ratio, associating higher levels of ER\u00ed2 with tumor relapse [138]. Yet despite their apparent statistical significance, the differences between ER\$1, ER\$2 and tumor relapse appeared fairly modest, which could be due to the relatively modest sample size or to the researchers' sole use of immunohistochemistry to detect the individual isoforms. Though the clinical significance of this particular study is somewhat questionable, an earlier prostate cancer study carried out by Leung et al. did reveal a correlation between the increased expression of ERB2 and ERB5 and enhanced metastasis and poor prognosis [139]. Clearly more studies are needed to better understand the prognostic value of the various ERB isoforms in breast cancer.

Regardless, the overwhelming consensus that ERβ is a positive prognostic marker is due largely to the fact that a significant number of ERβ+ patients respond well to endocrine therapy, such as tamoxifen. Several clinical studies have shown that patients who are ERα-negative or even triple negative (ERar/PR-/HER2-) but express high levels of ERB respond well to tamoxifen [111, 128, 130], whereas low levels of ERB correlate with resistance to tamoxifen treatment [140, 141]. In addition, other chemotherapeutic agents such as doxorubicin and cisplatin have been found to be more effective on breast cancer cell lines that express ER\$5 [142]. Aside from drug response, modification of ERB has recently been shown to indicate good prognosis as well. Specifically, a clinical study led by Valerie Speirs found that breast cancer patients expressing ERB phosphorylated at serine 105 had more favorable prognosis [143]. Though the mechanism behind this finding is unknown, it adds yet another level of complexity to an already complicated ERB narrative.

PR-positive breast cancer

ERα/PR+ breast cancers only account for about 2-7% of total breast cancer cases [81, 144-146], although there is controversy over whether or not such cancers actually exist. While many studies have supported the claim that ERα/PR+ is a distinct class of breast tumors [81, 94, 144-152], skeptics contend that since PR is an ER target gene, ER expression is a prerequisite for the expression of PR [153-155]. They maintain that the PR positivity in ERα-

negative tumors may simply reflect methodological errors in detecting PR and/or ERa, resulting in either a false-negative ERα result or a false-positive PR result [153, 154]. In fact, it is possible that tumors denoted as ERa-PR+ may actually have fairly low levels of $ER\alpha^{-}$ far below the sensitivity of the current assays [153]. Furthermore, results from Iwase et al. have indicated the possibility that the presence of ER α variants may exist in ER α -/PR+ tumors, specifically a variant with a deletion of exon 5 [156]. Though the presence of this variant in human breast tumors was confirmed in three other studies [157-159], this and other ERa variants may be difficult to detect via the traditional ERa assays used in immunohistochemistry, thus complicating the ERa status of certain mammary tumors. However, ERα aside, the PR promoter has been shown to be regulated by other transcription factors such as AP-1 and SP-1 [160-162]. In fact, an earlier study by Encarnación et al. indicated that while most of the ERα+ tumors converted to an ERα- phenotype, the PR status remained unchanged, further supporting the existence of an ERα⁻/PR⁺ clinical subtype and the possibility that PR can be regulated by signaling pathways other than ERα [163].

Although the expression of PR in breast cancer cells has been linked to both positive endocrine response and clinical outcome [82], it is unclear how PR affects the outcome in ERα-negative breast cancers. Multiple reports have indicated that ERa /PR breast tumors constitute a distinct clinicopathological group of cancers that results in outcomes worse than those that are $ER\alpha^+/PR^+$ or $ER\alpha^+/PR^-$, yet results in a better prognosis than double negative tumors (ERa-/ PR⁻) [144, 152, 164-166]. It is questionable whether this clinical subtype of tumor is responsive to endocrine therapies like tamoxifen. As noted earlier, PR has been shown to be an indicator of responsiveness to endocrine therapy in the presence of ERa, and thus it is conceivable that ERa⁻/PR⁺ tumors may also respond to tamoxifen. On the other hand, since current endocrine therapies are believed to target the ERα signaling pathway, it seems counterintuitive to attempt such therapeutic options on ERα⁻/PR⁺ cancer patients, particularly since the presence of PR in ERα+ tissues merely indicates a functional ERa, the presence of which is believed necessary for tamoxifen to have any

effect. Yet a recent study by Yang et al. found that patients with low-grade $ER\alpha/PR^+$ tumors did experience an overall survival benefit from adjuvant hormone therapy using tamoxifen; and conversely, no benefit was observed in patients with high-grade $ER\alpha/PR^+$ tumors [96]. Unfortunately, the scarcity of studies on this particular clinical subtype limits our ability to do a complete evaluation of the effectiveness of tamoxifen.

Theoretically, tumors that are ERα⁻/PR⁺ should be treatable with selective progesterone receptor modulators (SPRM), also referred to as antiprogestins. Anti-progestins such as mifepristone (RU-486) have been proposed as a new form of endocrine treatment or as an adjunct to the anti-estrogenic treatments for breast cancer. Multiple in vitro studies using cancer cell lines have indicated that low doses of anti-progestins can inhibit PR- and estrogen-mediated cell proliferation [167-169]. Contrary to these observations, other studies have demonstrated that at higher concentrations, mifepristone and other anti-progestins can actually stimulate proliferation of the ER+/PR+ breast cancer cells, T47D and MCF7 [170-172], indicating that the effect of anti-progestins on proliferation is dose-dependent and perhaps even dependent on the level of functional ERα. The few clinical studies that have tested the effectiveness of anti-progestins show limited to minimal efficacy in treating PR+ breast cancer [173-175]. Yet there is evidence that anti-progestins can augment the effects of anti-estrogens like tamoxifen [176-178]. Clearly, further clinical studies are necessary to determine if PR is a viable therapeutic target in ERa breast cancer.

AR-positive breast cancer

AR has been found in a significant number of $ER\alpha$ -negative tumors as well, with 22 to 49% of $ER\alpha$ -negative breast tumors expressing AR, depending on the clinical study [114, 118, 179, 180]. The presence of AR in $ER\alpha$ breast cancer is often associated with lower tumor grade, smaller tumor size, and significant increase in survival rates. Results inconsistent with these clinical data have been reported by some researchers who have found that androgenenhanced expression of AR in the ER/PR/HER2+ breast cancer cell line MDA-MB-453 increases cell proliferation [181, 182], which

can be inhibited by the AR antagonist bicalutamide [182]. Additionally, Richer et al. found that the AR inhibitor enzalutamide inhibited cell proliferation in ER α /AR $^+$ breast cancer cell lines [122]. Given that isoforms of AR were not taken into consideration in any of these studies, such contradictory findings between clinical and *in vitro* studies are not that surprising.

A fraction of triple-negative breast cancer cases (13-35%) appear to express AR [180, 183, 184]. Although some in vitro studies on triple-negative/AR+ cell lines have demonstrated an androgen-induced increase in cell proliferation [182, 185], most clinical studies find that the presence of AR in triple-negative tumors is correlated with a lower recurrence rate, fewer positive lymph node and distant metastases, lower histological grade, and higher overall survival rate compared to triple-negative tumors that are AR-negative [180, 183, 184, 186]. These data support the notion that AR can be used as a positive prognostic factor, not only in ERα-negative but also triple-negative breast cancers.

In addition to its prospective use as a prognostic biomarker for breast cancer, additional studies have indicated that AR may also serve as a potential therapeutic target for those breast cancer patients who traditionally have had very few treatment options, such as those whose tumors are ER-/PR-/AR+. Hardin and colleagues tested the effectiveness of the androgen dehydroepiandrosterone sulfate (DHEAS) on ER-/ PR-/AR+ breast cancer cell lines and found that DHEAS stimulation of AR hampered cell growth by enhancing apoptosis [187]. Conflicting studies by Ni et al. and Arce-Salinas et al. showed that inhibiting AR with the antagonist bicalutamide led to growth inhibition and enhanced cell death [182, 188]. Interestingly, AR expression seems to be correlated with the overexpression of the oncogene HER2 via a complisignaling cascade that involves upregulation of HER2 by AR and transcription factor β-catenin [182]. In addition, expression of the constitutively active AR variant AR-V7 in both ERα breast cancer cell lines (e.g. MDA-MB-453) and ERα primary tissues is actually enhanced by the AR antagonist enzalutamide, which in turn increases cell growth and, as with more advanced stages of prostate cancer, could result in ADT resistance [71]. Clearly the

use of AR as a therapeutic target is wrought with complications, as both agonists and antagonists of AR have succeeded in either suppressing or promoting tumor progression, and success appears to be highly dependent on AR isoform expression and ER α levels.

Discussion and conclusion

Triple negative breast cancer (i.e. ER-/PR-/ HER2-) is virtually impossible to treat with the established endocrine-based therapies, as such therapies were originally intended for cancers that are ERα-positive. However, our review illustrates how confining the triple negative designation can be, potentially preventing clinicians from considering other factors that play a role in breast cancer progression. Data obtained from basic and translational research studies indicate that there are indeed many other proteins involved in breast cancer development and progression; but despite these findings, most clinical pathologists continue to follow the standard practice of categorizing breast tumors using only the three established markers- ERα, PR, and HER2. This apparent disconnect between the bench and the clinic is alarming when one considers that 10-15% of breast cancers are diagnosed as triple negative [2, 3] and that even patients diagnosed as ERapositive do not all respond equally well to antiestrogen treatment, which further demonstrates the complexity of breast cancer and emphasizes the need for more therapeutic targets.

The goal of this review was to determine if other steroid hormone receptors- most notably ERB and AR- should also be routinely analyzed and used as additional targets for breast cancer treatment and prognosis. Although a significant number of studies exist describing the presence of ERB, AR, and PR in breast cancer cells or tissues, obtaining truly accurate expression levels from the literature proved difficult as all three of these receptors exist in at least two isoform states. Most studies did not account for this variability. Therefore, it is probable that many isoforms went undetected, which in turn could have led to an underestimate of a receptor's actual expression level. Yet, quantitative inaccuracies aside, we still found very strong evidence that each of these receptors influences tumor progression, either negatively or positively, depending upon which isoforms are present and the level of ER α expression. For example, ERα⁺/PR⁺ breast tumors are generally found to be more responsive to endocrine treatment than $ER\alpha^+/PR^-$ tumors [85, 91, 93-96]. and yet a higher expression level of the PR-A isoform compared to PR-B has been associated with anti-estrogen resistance and subsequently poorer prognosis [102]. The isoforms of ERB are capable of forming heterodimers with $ER\alpha$, inhibiting $ER\alpha$ activity and consequently resulting in tumor suppression. However, other studies have shown a positive correlation between tumor progression and increased levels of ERβ2/cx and ERβ5 isoforms relative to ERB1 [135, 138, 139]. The expression of certain isoforms, such as ER\$5, has also been correlated with positive drug response [142]. Unfortunately, due to the relative scarcity of studies analyzing AR in breast tumors, few clinically significant associations were found between a specific AR isoform and breast tumor progression. However, certain splice variants such as the ligand-independent isoform AR-V7 have been shown to impact tumor development and progression in prostate cancer [67, 68], and a recent study by the Tilley lab strongly suggest that it may have the same impact on breast cancer progression [71]. Additionally, the available data do show that the presence of AR is correlated with good prognosis for both ERα-positive and ERαnegative breast cancer, indicating that antiandrogen therapy might be a viable option in certain breast cancers as it is in prostate cancer.

We acknowledge that more basic research is necessary to elucidate the molecular mechanisms behind the steroid hormone receptors' effects on breast cancer development and progression. In particular, more studies are needed to better understand how ERB, PR, and AR influence cell proliferation, apoptosis, and metastasis. However, in the interest of saving lives, we feel that the existing data justify collecting more clinical data on a more massive scale. We contend that all breast tumor biopsies should be analyzed not only for ERα, HER2, and overall PR expression but also for each isoform of ERα, PR, ERβ, and AR. Obviously this is an ambitious undertaking that would require developing new techniques and improving current detection methods for each isoform. In

addition, subsequent large-scale analysis of the data will be necessary to determine if consistent patterns emerge linking the various expression levels of these markers and their isoforms with particular grades and stages of breast cancer.

More and more clinical labs are trending towards molecular diagnostic procedures such as next generation sequencing (NGS), RNA sequencing, and high-throughput qRT-PCR, all of which will allow scientists to analyze more genetic markers in a relatively short period of time. Additionally, the isolation and analysis of circulating cell-free DNA (ccfDNA) could allow for initial prognosis using very little sample. In fact, several clinical and translational studies have already shown how NGS and ccfDNA analysis can aid in breast cancer prognosis and tumor classification [189-193]. However, techniques such as NGS are best suited for patients who may have a genetic predisposition for a particular cancer (e.g. BRCA1 and BRCA2 in some familial breast cancers); and though RNA sequencing and qRT-PCR do give information regarding transcriptional expression, it is at the protein level that phenotypes are often determined.

Therefore, although the utilization of molecular techniques will enable clinicians to obtain more information about a patient's tumor, these techniques should always be used in conjunction with protein expression analyses-particularly since not all of the steroid receptor isoforms are due to splicing variations but instead are generated post-translationally. In addition, post-translational modifications such as phosphorylation and glycosylation cannot be detected at the DNA or RNA level but only by analyzing the protein expression profiles of tumors. Finally, the heterogeneity of the cell population within an individual tumor must be taken into account when analyzing all the data this new technology will generate. To date, the most common procedures for studying protein expression are immunohistochemistry and western blotting. Though immunohistochemistry provides an important visual of protein expression variations between tissues and even between cells within the tumor, and western blotting is a more stringent and exact method for detecting isoforms of varying molecular weights, both depend on antibody-binding,

which is an indirect method of protein detection. Recent advances in multiplexed protein analysis include mass spectrometry immunohistochemistry (MSIHC), in which metal tags of varying masses are used to label antibodies. thus allowing for the simultaneous detection of up to 100 different protein targets [194]. Such a technique has been used successfully in analyzing various markers such as ERa, PR, and HER2 in breast tumor samples [195]. Again, however, such a technique still relies on antibody recognition and binding. Another technique that involves rapid protein isolation and sequencing on a large-scale would need to be developed in future to directly identify the various steroid receptor isoforms expressed in a given tumor. Ultimately, we envision a multipronged approach to breast cancer analysis that will allow detection of a variety of markers, including the many isoforms of ERa, ERB, PR, and AR, coupled with the pathology of the tumor. The overall goal would be to provide each patient with a more personalized and accurate prognosis and more effective treatment options, and maybe even render obsolete the term "triple negative breast cancer".

Acknowledgements

We thank Esmeralda Ponce, Mariah Alejo, and Rubi Garcia for their support. This research did not receive any specific grant from funding agencies in the public, commercial, or not-forprofit sectors.

Disclosure of conflict of interest

None.

Abbreviations

ER, estrogen receptor; PR, progesterone receptor; AR, androgen receptor; HER2, human epidermal growth factor receptor type 2; GR, glucocorticoid receptor; GPR-30, G-protein coupled receptor-30; DBD, DNA binding domain; LBD, ligand binding domain; HRE, hormone response element; NTD, N-terminal domain; SERMs, selective ER modulators; SERDs, selective ER down-regulators.

Address correspondence to: Dr. Mary B Sevigny, Department of Natural Sciences and Mathematics, Dominican University of California, 50 Acacia Avenue, San Rafael, CA 94901, USA. Tel: 415-458-

3743; Fax: 415-482-1972; E-mail: mary.sevigny@dominican.edu

References

- American Cancer Society. Cancer facts & figures 2016. Atlanta American Cancer Society; 2016.
- [2] Dawood S. Triple-negative breast cancer: epidemiology and management options. Drugs 2010; 70: 2247-2258.
- [3] Foulkes WD, Smith IE and Reis-Filho JS. Triplenegative breast cancer. N Engl J Med 2010; 363: 1938-1948.
- [4] Beato M and Klug J. Steroid hormone receptors: an update. Hum Reprod Update 2000; 6: 225-236.
- [5] MacGregor JI and Jordan VC. Basic guide to the mechanisms of antiestrogen action. Pharmacol Rev 1998; 50: 151-196.
- [6] Prossnitz ER, Arterburn JB and Sklar LA. GPR30: a G protein-coupled receptor for estrogen. Mol Cell Endocrinol 2007; 265-266: 138-142.
- [7] Stossi F, Madak-Erdogan Z and Katzenellenbogen BS. Estrogen receptor alpha represses transcription of early target genes via p300 and CtBP1. Mol Cell Biol 2009; 29: 1749-1759.
- [8] Frasor J, Danes JM, Komm B, Chang KC, Lyttle CR and Katzenellenbogen BS. Profiling of estrogen up- and down-regulated gene expression in human breast cancer cells: insights into gene networks and pathways underlying estrogenic control of proliferation and cell phenotype. Endocrinology 2003; 144: 4562-4574.
- [9] Matthews J and Gustafsson JA. Estrogen signaling: a subtle balance between ER alpha and ER beta. Mol Interv 2003; 3: 281-292.
- [10] Huang J, Li X, Yi P, Hilf R, Bambara RA and Muyan M. Targeting estrogen responsive elements (EREs): design of potent transactivators for ERE-containing genes. Mol Cell Endocrinol 2004; 218: 65-78.
- [11] Huang P, Chandra V and Rastinejad F. Structural overview of the nuclear receptor superfamily: insights into physiology and therapeutics. Annu Rev Physiol 2010; 72: 247-272.
- [12] Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, Tujague M, Strom A, Treuter E, Warner M and Gustafsson JA. Estrogen receptors: how do they signal and what are their targets. Physiol Rev 2007; 87: 905-931.
- [13] Berry M, Metzger D and Chambon P. Role of the two activating domains of the oestrogen receptor in the cell-type and promoter-context dependent agonistic activity of the anti-oestro-

- gen 4-hydroxytamoxifen. EMBO J 1990; 9: 2811-2818.
- [14] O'Malley BW and Khan S. Elwood V. Jensen (1920-2012): father of the nuclear receptors. Proc Natl Acad Sci U S A 2013; 110: 3707-3708.
- [15] Jensen EV and Jordan VC. The estrogen receptor: a model for molecular medicine. Clin Cancer Res 2003; 9: 1980-1989.
- [16] Carmeci C, Thompson DA, Ring HZ, Francke U and Weigel RJ. Identification of a Gene (GPR30) with homology to the G-protein-coupled receptor superfamily associated with estrogen receptor expression in breast cancer. Genomics 1997; 45: 607-617.
- [17] Filardo EJ, Quinn JA, Frackelton AR and Bland KI. Estrogen action via the G protein-coupled receptor, GPR30: stimulation of adenylyl cyclase and cAMP-mediated attenuation of the epidermal growth factor receptor-to-MAPK signaling axis. Mol Endocrinol 2002; 16: 70-84.
- [18] Filardo EJ and Thomas P. GPR30: a seventransmembrane-spanning estrogen receptor that triggers EGF release. Trends Endocrinol Metab 2005; 16: 362-367.
- [19] Hewitt SC and Korach KS. Estrogen receptors: structure, mechanisms and function. Rev Endocr Metab Disord 2002; 3: 193-200.
- [20] Nilsson S, Makela S, Treuter E, Tujague M, Thomsen J, Andersson G, Enmark E, Pettersson K, Warner M and Gustafsson JA. Mechanisms of estrogen action. Physiol Rev 2001; 81: 1535-1565.
- [21] Albanito L, Lappano R, Madeo A, Chimento A, Prossnitz ER, Cappello AR, Dolce V, Abonante S, Pezzi V and Maggiolini M. G-protein-coupled receptor 30 and estrogen receptor-alpha are involved in the proliferative effects induced by atrazine in ovarian cancer cells. Environ Health Perspect 2008; 116: 1648-1655.
- [22] Prossnitz ER, Arterburn JB, Smith HO, Oprea TI, Sklar LA and Hathaway HJ. Estrogen signaling through the transmembrane G protein-coupled receptor GPR30. Annu Rev Physiol 2008; 70: 165-190.
- [23] Prossnitz ER, Oprea TI, Sklar LA and Arterburn JB. The ins and outs of GPR30: a transmembrane estrogen receptor. J Steroid Biochem Mol Biol 2008; 109: 350-353.
- [24] Speirs V, Skliris GP, Burdall SE and Carder PJ. Distinct expression patterns of ER alpha and ER beta in normal human mammary gland. J Clin Pathol 2002; 55: 371-374.
- [25] Mehta RG, Hawthorne M, Mehta RR, Torres KE, Peng X, McCormick DL and Kopelovich L. Differential roles of ERalpha and ERbeta in normal and neoplastic development in the mouse mammary gland. PLoS One 2014; 9: e113175.

- [26] Platet N, Cathiard AM, Gleizes M and Garcia M. Estrogens and their receptors in breast cancer progression: a dual role in cancer proliferation and invasion. Crit Rev Oncol Hematol 2004; 51: 55-67.
- [27] Saji S, Jensen EV, Nilsson S, Rylander T, Warner M and Gustafsson JA. Estrogen receptors alpha and beta in the rodent mammary gland. Proc Natl Acad Sci U S A 2000; 97: 337-342.
- [28] Stope MB, Popp SL, Knabbe C and Buck MB. Estrogen receptor alpha attenuates transforming growth factor-beta signaling in breast cancer cells independent from agonistic and antagonistic ligands. Breast Cancer Res Treat 2010; 120: 357-367.
- [29] Babiker FA, De Windt LJ, van Eickels M, Grohe C, Meyer R and Doevendans PA. Estrogenic hormone action in the heart: regulatory network and function. Cardiovasc Res 2002; 53: 709-719.
- [30] Halon A, Materna V, Drag-Zalesinska M, Nowak-Markwitz E, Gansukh T, Donizy P, Spaczynski M, Zabel M, Dietel M, Lage H and Surowiak P. Estrogen receptor alpha expression in ovarian cancer predicts longer overall survival. Pathol Oncol Res 2011; 17: 511-518.
- [31] Mylonas I, Jeschke U, Shabani N, Kuhn C, Balle A, Kriegel S, Kupka MS and Friese K. Immunohistochemical analysis of estrogen receptor alpha, estrogen receptor beta and progesterone receptor in normal human endometrium. Acta Histochemica 2004; 106: 245-252.
- [32] Taylor AH and Al-Azzawi F. Immunolocalisation of oestrogen receptor beta in human tissues. J Mol Endocrinol 2000; 24: 145-155.
- [33] Flouriot G, Brand H, Denger S, Metivier R, Kos M, Reid G, Sonntag-Buck V and Gannon F. Identification of a new isoform of the human estrogen receptor-alpha (hER-alpha) that is encoded by distinct transcripts and that is able to repress hER-alpha activation function 1. EMBO J 2000; 19: 4688-4700.
- [34] Wang Z, Zhang X, Shen P, Loggie BW, Chang Y and Deuel TF. Identification, cloning, and expression of human estrogen receptor-alpha36, a novel variant of human estrogen receptor-alpha66. Biochem Biophys Res Commun 2005; 336: 1023-1027.
- [35] Wang Z, Zhang X, Shen P, Loggie BW, Chang Y and Deuel TF. A variant of estrogen receptor-{alpha}, hER-{alpha}36: transduction of estrogen- and antiestrogen-dependent membrane-initiated mitogenic signaling. Proc Natl Acad Sci U S A 2006; 103: 9063-9068.
- [36] Lin AH, Li RW, Ho EY, Leung GP, Leung SW, Vanhoutte PM and Man RY. Differential ligand binding affinities of human estrogen receptoralpha isoforms. PLoS One 2013; 8: e63199.

- [37] Leung YK, Mak P, Hassan S and Ho SM. Estrogen receptor (ER)-β isoforms: a key to understanding ER-β signaling. Proc Natl Acad Sci U S A 2006; 103: 13162-13167.
- [38] Ogawa S, Inoue S, Watanabe T, Orimo A, Hosoi T, Ouchi Y and Muramatsu M. Molecular cloning and characterization of human estrogen receptor betacx: a potential inhibitor ofestrogen action in human. Nucleic Acids Res 1998; 26: 3505-3512.
- [39] Omoto Y, Eguchi H, Yamamoto-Yamaguchi Y and Hayashi S. Estrogen receptor (ER) beta1 and ERbetacx/beta2 inhibit ERalpha function differently in breast cancer cell line MCF7. Oncogene 2003; 22: 5011-5020.
- [40] Poola I, Abraham J, Baldwin K, Saunders A and Bhatnagar R. Estrogen receptors beta4 and beta5 are full length functionally distinct ERbeta isoforms: cloning from human ovary and functional characterization. Endocrine 2005; 27: 227-238.
- [41] Moore JT, McKee DD, Slentz-Kesler K, Moore LB, Jones SA, Horne EL, Su JL, Kliewer SA, Lehmann JM and Willson TM. Cloning and characterization of human estrogen receptor beta isoforms. Biochem Biophys Res Commun 1998; 247: 75-78.
- [42] Skliris GP, Leygue E, Curtis-Snell L, Watson PH and Murphy LC. Expression of oestrogen receptor-β in oestrogen receptor-α negative human breast tumours. Br J Cancer 2006; 95: 616-626.
- [43] Leygue E and Murphy LC. A bi-faceted role of estrogen receptor beta in breast cancer. Endocr Relat Cancer 2013; 20: R127-139.
- [44] Carroll JS and Brown M. Estrogen receptor target gene: an evolving concept. Mol Endocrinol 2006; 20: 1707-1714.
- [45] Carroll JS, Meyer CA, Song J, Li W, Geistlinger TR, Eeckhoute J, Brodsky AS, Keeton EK, Fertuck KC, Hall GF, Wang Q, Bekiranov S, Sementchenko V, Fox EA, Silver PA, Gingeras TR, Liu XS and Brown M. Genome-wide analysis of estrogen receptor binding sites. Nat Genet 2006; 38: 1289-1297.
- [46] Kastner P, Krust A, Turcotte B, Stropp U, Tora L, Gronemeyer H and Chambon P. Two distinct estrogen-regulated promoters generate transcripts encoding the two functionally different human progesterone receptor forms A and B. EMBO J 1990; 9: 1603-1614.
- [47] Conneely OM and Lydon JP. Progesterone receptors in reproduction: functional impact of the A and B isoforms. Steroids 2000; 65: 571-577
- [48] Lydon JP, DeMayo FJ, Funk CR, Mani SK, Hughes AR, Montgomery CA Jr, Shyamala G, Conneely OM, O'Malley BW. Mice lacking progesterone receptor exhibit pleiotropic repro-

- ductive abnormalities. Genes Dev 1995; 9: 2266-2278.
- [49] Conneely OM, Mulac-Jericevic B and Lydon JP. Progesterone-dependent regulation of female reproductive activity by two distinct progesterone receptor isoforms. Steroids 2003; 68: 771-778.
- [50] Mulac-Jericevic B, Lydon JP, DeMayo FJ and Conneely OM. Defective mammary gland morphogenesis in mice lacking the progesterone receptor B isoform. Proc Natl Acad Sci U S A 2003; 100: 9744-9749.
- [51] Mulac-Jericevic B, Mullinax RA, DeMayo FJ, Lydon JP and Conneely OM. Subgroup of reproductive functions of progesterone mediated by progesterone receptor-B isoform. Science (New York, N.Y.) 2000; 289: 1751-1754.
- [52] Soyal S, Ismail PM, Li J, Mulac-Jericevic B, Conneely OM and Lydon JP. Progesterone's role in mammary gland development and tumorigenesis as disclosed by experimental mouse genetics. Breast Cancer Res 2002; 4: 191-196.
- [53] Humphreys RC, Lydon JP, O'Malley BW and Rosen JM. Use of PRKO mice to study the role of progesterone in mammary gland development. J Mammary Gland Biol Neoplasia 1997; 2: 343-354.
- [54] Humphreys RC, Lydon J, O'Malley BW and Rosen JM. Mammary gland development is mediated by both stromal and epithelial progesterone receptors. Mol Endocrinol 1997; 11: 801-811.
- [55] Richer JK, Jacobsen BM, Manning NG, Abel MG, Wolf DM and Horwitz KB. Differential gene regulation by the two progesterone receptor isoforms in human breast cancer cells. J Biol Chem 2002; 277: 5209-5218.
- [56] Boonyaratanakornkit V, McGowan E, Sherman L, Mancini MA, Cheskis BJ and Edwards DP. The role of extranuclear signaling actions of progesterone receptor in mediating progesterone regulation of gene expression and the cell cycle. Mol Endocrinol 2007; 21: 359-375.
- [57] Graham JD, Yager ML, Hill HD, Byth K, O'Neill GM and Clarke CL. Altered progesterone receptor isoform expression remodels progestin responsiveness of breast cancer cells. Mol Endocrinol 2005; 19: 2713-2735.
- [58] Hopp TA, Weiss HL, Hilsenbeck SG, Cui Y, Allred DC, Horwitz KB and Fuqua SA. Breast cancer patients with progesterone receptor PR-A-rich tumors have poorer disease-free survival rates. Clin Cancer Res 2004; 10: 2751-2760.
- [59] Dimitrakakis C and Bondy C. Androgens and the breast. Breast Cancer Res 2009; 11: 212.
- [60] Gelmann EP. Molecular biology of the androgen receptor. J Clin Oncol 2002; 20: 3001-3015.

- [61] Ruizeveld de Winter JA, Trapman J, Vermey M, Mulder E, Zegers ND and van der Kwast TH. Androgen receptor expression in human tissues: an immunohistochemical study. J Histochem Cytochem 1991; 39: 927-936.
- [62] Roy AK, Lavrovsky Y, Song CS, Chen S, Jung MH, Velu NK, Bi BY and Chatterjee B. Regulation of androgen action. Vitam Horm 1999; 55: 309-352.
- [63] Lu NZ, Wardell SE, Burnstein KL, Defranco D, Fuller PJ, Giguere V, Hochberg RB, McKay L, Renoir JM, Weigel NL, Wilson EM, McDonnell DP and Cidlowski JA. International Union of Pharmacology. LXV. The pharmacology and classification of the nuclear receptor superfamily: glucocorticoid, mineralocorticoid, progesterone, and androgen receptors. Pharmacol Rev 2006; 58: 782-797.
- [64] Poletti A. The polyglutamine tract of androgen receptor: from functions to dysfunctions in motor neurons. Front Neuroendocrinol 2004; 25: 1-26.
- [65] Wilson CM and McPhaul MJ. A and B forms of the androgen receptor are present in human genital skin fibroblasts. Proc Natl Acad Sci U S A 1994; 91: 1234-1238.
- [66] Mudryj M and Tepper CG. On the origins of the androgen receptor low molecular weight species. Horm Cancer 2013; 4: 259-269.
- [67] Guo Z, Yang X, Sun F, Jiang R, Linn DE, Chen H, Chen H, Kong X, Melamed J, Tepper CG, Kung HJ, Brodie AM, Edwards J and Qiu Y. A novel androgen receptor splice variant is up-regulated during prostate cancer progression and promotes androgen depletion-resistant growth. Cancer Res 2009; 69: 2305-2313.
- [68] Hu R, Dunn TA, Wei S, Isharwal S, Veltri RW, Humphreys E, Han M, Partin AW, Vessella RL, Isaacs WB, Bova GS and Luo J. Ligand-independent androgen receptor variants derived from splicing of cryptic exons signify hormonerefractory prostate cancer. Cancer Res 2009; 69: 16-22.
- [69] Antonarakis ES, Lu C, Wang H, Luber B, Nakazawa M, Roeser JC, Chen Y, Mohammad TA, Chen Y, Fedor HL, Lotan TL, Zheng Q, De Marzo AM, Isaacs JT, Isaacs WB, Nadal R, Paller CJ, Denmeade SR, Carducci MA, Eisenberger MA and Luo J. AR-V7 and resistance to enzalutamide and abiraterone in prostate cancer. N Engl J Med 2014; 371: 1028-1038.
- [70] Li Y, Chan SC, Brand LJ, Hwang TH, Silverstein KA and Dehm SM. Androgen receptor splice variants mediate enzalutamide resistance in castration-resistant prostate cancer cell lines. Cancer Res 2013; 73: 483-489.
- [71] Hickey TE, Irvine CM, Dvinge H, Tarulli GA, Hanson AR, Ryan NK, Pickering MA, Birrell SN, Hu DG, Mackenzie PI, Russell R, Caldas C, Raj GV,

- Dehm SM, Plymate SR, Bradley RK, Tilley WD and Selth LA. Expression of androgen receptor splice variants in clinical breast cancers. Oncotarget 2015; 6: 44728-44744.
- [72] Zhu X, Daffada AA, Chan CM and Dowsett M. Identification of an exon 3 deletion splice variant androgen receptor mRNA in human breast cancer. Int J Cancer 1997; 72: 574-580.
- [73] Quigley CA, Evans BA, Simental JA, Marschke KB, Sar M, Lubahn DB, Davies P, Hughes IA, Wilson EM and French FS. Complete androgen insensitivity due to deletion of exon C of the androgen receptor gene highlights the functional importance of the second zinc finger of the androgen receptor in vivo. Mol Endocrinol 1992; 6: 1103-1112.
- [74] Ali S and Coombes RC. Estrogen receptor alpha in human breast cancer: occurrence and significance. J Mammary Gland Biol Neoplasia 2000; 5: 271-281.
- [75] Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S and Gustafsson JA. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. Endocrinology 1997; 138: 863-870.
- [76] Heldring N, Pawson T, McDonnell D, Treuter E, Gustafsson JA and Pike AC. Structural insights into corepressor recognition by antagonistbound estrogen receptors. J Biol Chem 2007; 282: 10449-10455.
- [77] Yeh WL, Shioda K, Coser KR, Rivizzigno D, Mc-Sweeney KR and Shioda T. Fulvestrant-induced cell death and proteasomal degradation of estrogen receptor alpha protein in MCF-7 cells require the CSK c-Src tyrosine kinase. PLoS One 2013; 8: e60889.
- [78] Long X and Nephew KP. Fulvestrant (ICI 182,780)-dependent interacting proteins mediate immobilization and degradation of estrogen receptor-alpha. J Biol Chem 2006; 281: 9607-9615.
- [79] Kocanova S, Mazaheri M, Caze-Subra S and Bystricky K. Ligands specify estrogen receptor alpha nuclear localization and degradation. BMC Cell Biol 2010; 11: 98.
- [80] Arpino G, Weiss H, Lee AV, Schiff R, Placido SD, Osborne CK and Elledge RM. Estrogen receptor-positive, progesterone receptor-negative breast cancer: association with growth factor receptor expression and tamoxifen resistance. J Natl Cancer Inst 2005; 97: 1254-1261.
- [81] Chu KC, Anderson WF, Fritz A, Ries LA and Brawley OW. Frequency distributions of breast cancer characteristics classified by estrogen receptor and progesterone receptor status for eight racial/ethnic groups. Cancer 2001; 92: 37-45.

- [82] Cui X, Schiff R, Arpino G, Osborne CK and Lee AV. Biology of progesterone receptor loss in breast cancer and its implications for endocrine therapy. J Clin Oncol 2005; 23: 7721-7735
- [83] Dunnwald LK, Rossing MA and Li Cl. Hormone receptor status, tumor characteristics, and prognosis: a prospective cohort of breast cancer patients. Breast Cancer Res 2007; 9: R6.
- [84] Reiner A, Neumeister B, Spona J, Reiner G, Schemper M and Jakesz R. Immunocytochemical localization of estrogen and progesterone receptor and prognosis in human primary breast cancer. Cancer Res 1990; 50: 7057-7061.
- [85] Bardou VJ, Arpino G, Elledge RM, Osborne CK and Clark GM. Progesterone receptor status significantly improves outcome prediction over estrogen receptor status alone for adjuvant endocrine therapy in two large breast cancer databases. J Clin Oncol 2003; 21: 1973-1979.
- [86] Pichon MF, Pallud C, Brunet M and Milgrom E. Relationship of presence of progesterone receptors to prognosis in early breast cancer. Cancer Res 1980; 40: 3357-3360.
- [87] Salmen J, Neugebauer J, Fasching PA, Haeberle L, Huober J, Wockel A, Rauh C, Schuetz F, Weissenbacher T, Kost B, Stickeler E, Klar M, Orlowska-Volk M, Windfuhr-Blum M, Heil J, Rom J, Sohn C, Fehm T, Mohrmann S, Loebberg CR, Hein A, Schulz-Wendtland R, Hartkopf AD, Brucker SY, Wallwiener D, Friese K, Hartmann A, Beckmann MW, Janni W and Rack B. Pooled analysis of the prognostic relevance of progesterone receptor status in five German cohort studies. Breast Cancer Res Treat 2014; 148: 143-151.
- [88] Elledge RM, Green S, Pugh R, Allred DC, Clark GM, Hill J, Ravdin P, Martino S and Osborne CK. Estrogen receptor (ER) and progesterone receptor (PgR), by ligand-binding assay compared with ER, PgR and pS2, by immuno-histochemistry in predicting response to tamoxifen in metastatic breast cancer: a Southwest Oncology Group Study. Int J Cancer 2000; 89: 111-117.
- [89] McGuire WL. Steroid receptors in human breast cancer. Cancer Res 1978; 38: 4289-4291.
- [90] Osborne CK and Schiff R. Mechanisms of endocrine resistance in breast cancer. Annu Rev Med 2011; 62: 233-247.
- [91] Osborne CK, Yochmowitz MG, Knight WA and McGuire WL. The value of estrogen and progesterone receptors in the treatment of breast cancer. Cancer 1980; 46: 2884-2888.
- [92] Horwitz KB, Costlow ME and McGuire WL. MCF-7; a human breast cancer cell line with estrogen, androgen, progesterone, and gluco-

- corticoid receptors. Steroids 1975; 26: 785-795.
- [93] Bae SY, Kim S, Lee JH, Lee HC, Lee SK, Kil WH, Kim SW, Lee JE and Nam SJ. Poor prognosis of single hormone receptor- positive breast cancer: similar outcome as triple-negative breast cancer. BMC Cancer 2015; 15: 138.
- [94] Ferno M, Stal O, Baldetorp B, Hatschek T, Kallstrom AC, Malmstrom P, Nordenskjold B and Ryden S. Results of two or five years of adjuvant tamoxifen correlated to steroid receptor and S-phase levels. South Sweden Breast Cancer Group, and South-East Sweden Breast Cancer Group. Breast Cancer Res Treat 2000; 59: 69-76.
- [95] Purdie CA, Quinlan P, Jordan LB, Ashfield A, Ogston S, Dewar JA and Thompson AM. Progesterone receptor expression is an independent prognostic variable in early breast cancer: a population-based study. Br J Cancer 2014; 110: 565-572.
- [96] Yang LH, Tseng HS, Lin C, Chen LS, Chen ST, Kuo SJ and Chen DR. Survival benefit of tamoxifen in estrogen receptor-negative and progesterone receptor-positive low grade breast cancer patients. J Breast Cancer 2012; 15: 288-295.
- [97] Forouzanfar MH, Foreman KJ, Delossantos AM, Lozano R, Lopez AD, Murray CJ and Naghavi M. Breast and cervical cancer in 187 countries between 1980 and 2010: a systematic analysis. Lancet 2011; 378: 1461-1484.
- [98] Stendahl M, Ryden L, Nordenskjold B, Jonsson PE, Landberg G and Jirstrom K. High progesterone receptor expression correlates to the effect of adjuvant tamoxifen in premenopausal breast cancer patients. Clin Cancer Res 2006; 12: 4614-4618.
- [99] Clark GM, McGuire WL, Hubay CA, Pearson OH and Marshall JS. Progesterone receptors as a prognostic factor in Stage II breast cancer. N Engl J Med 1983; 309: 1343-1347.
- [100] Rutqvist LE, Cedermark B, Fornander T, Glas U, Johansson H, Nordenskjöld B, Rotstein S, Skoog L, Somell A, Theve T, et al. The relationship between hormone receptor content and the effect of adjuvant tamoxifen in operable breast cancer. J Clin Oncol 1989; 7: 1474-1484.
- [101] Bamberger AM, Milde-Langosch K, Schulte HM and Löning T. Progesterone receptor isoforms, PR-B and PR-A, in breast cancer: correlations with clinicopathologic tumor parameters and expression of AP-1 factors. Horm Res 2000; 54: 32-37.
- [102] Khan JA, Bellance C, Guiochon-Mantel A, Lombès M and Loosfelt H. Differential regulation of breast cancer-associated genes by progesterone receptor isoforms PRA and PRB in a new

- bi-inducible breast cancer cell line. PLoS One 2012; 7: e45993.
- [103] Allred DC, Harvey JM, Berardo M and Clark GM. Prognostic and predictive factors in breast cancer by immunohistochemical analysis. Mod Pathol 1998; 11: 155-168.
- [104] Gasparini G, Pozza F, Dittadi R, Meli S, Cazzavillan S and Bevilacqua P. Progesterone receptor determined by immunocytochemical and biochemical methods in human breast cancer. J Cancer Res Clin Oncol 1992; 118: 557-563.
- [105] Mote PA, Johnston JF, Manninen T, Tuohimaa P and Clarke CL. Detection of progesterone receptor forms A and B by immunohistochemical analysis. J Clin Pathol 2001; 54: 624-630.
- [106] Seymour L, Meyer K, Esser J, MacPhail AP, Behr A and Bezwoda WR. Estimation of PR and ER by immunocytochemistry in breast cancer. Comparison with radioligand binding methods. Am J Clin Pathol 1990; 94: S35-40.
- [107] Helguero LA, Faulds MH, Gustafsson JA and Haldosen LA. Estrogen receptors alfa (ERalpha) and beta (ERbeta) differentially regulate proliferation and apoptosis of the normal murine mammary epithelial cell line HC11. Oncogene 2005; 24: 6605-6616.
- [108] Strom A, Hartman J, Foster JS, Kietz S, Wimalasena J and Gustafsson JA. Estrogen receptor beta inhibits 17beta-estradiol-stimulated proliferation of the breast cancer cell line T47D. Proc Natl Acad Sci U S A 2004; 101: 1566-1571.
- [109] Peng B, Lu B, Leygue E and Murphy LC. Putative functional characteristics of human estrogen receptor-beta isoforms. J Mol Endocrinol 2003; 30: 13-29.
- [110] Wu X, Subramaniam M, Grygo SB, Sun Z, Negron V, Lingle WL, Goetz MP, Ingle JN, Spelsberg TC and Hawse JR. Estrogen receptor-beta sensitizes breast cancer cells to the anti-estrogenic actions of endoxifen. Breast Cancer Res 2011; 13: R27.
- [111] Gruvberger-Saal SK, Bendahl PO, Saal LH, Laakso M, Hegardt C, Eden P, Peterson C, Malmstrom P, Isola J, Borg A and Ferno M. Estrogen receptor beta expression is associated with tamoxifen response in ERalpha-negative breast carcinoma. Clin Cancer Res 2007; 13: 1987-1994.
- [112] Balk SP and Knudsen KE. AR, the cell cycle, and prostate cancer. Nucl Recept Signal 2008; 6: e001.
- [113] Moinfar F, Okcu M, Tsybrovskyy O, Regitnig P, Lax SF, Weybora W, Ratschek M, Tavassoli FA and Denk H. Androgen receptors frequently are expressed in breast carcinomas: potential relevance to new therapeutic strategies. Cancer 2003; 98: 703-711.

- [114] Hu R, Dawood S, Holmes MD, Collins LC, Schnitt SJ, Cole K, Marotti JD, Hankinson SE, Colditz GA and Tamimi RM. Androgen receptor expression and breast cancer survival in postmenopausal women. Clin Cancer Res 2011; 17: 1867-1874.
- [115] Agrawal AK, Jelen M, Grzebieniak Z, Zukrowski P, Rudnicki J and Nienartowicz E. Androgen receptors as a prognostic and predictive factor in breast cancer. Folia Histochem Cytobiol 2008; 46: 269-276.
- [116] Qu Q, Mao Y, Fei XC and Shen KW. The impact of androgen receptor expression on breast cancer survival: a retrospective study and meta-analysis. PLoS One 2013; 8: e82650.
- [117] Bryan RM, Mercer RJ, Bennett RC, Rennie GC, Lie TH and Morgan FJ. Androgen receptors in breast cancer. Cancer 1984; 54: 2436-2440.
- [118] Higa GM and Fell RG. Sex hormone receptor repertoire in breast cancer. Int J Breast Cancer 2013; 2013: 284036.
- [119] Rizza P, Barone I, Zito D, Giordano F, Lanzino M, De Amicis F, Mauro L, Sisci D, Catalano S, Dahlman Wright K, Gustafsson JA and Ando S. Estrogen receptor beta as a novel target of androgen receptor action in breast cancer cell lines. Breast Cancer Res 2014; 16: R21.
- [120] Paliouras M and Diamandis EP. Androgens act synergistically to enhance estrogen-induced upregulation of human tissue kallikreins 10, 11, and 14 in breast cancer cells via a membrane bound androgen receptor. Mol Oncol 2008; 1: 413-424.
- [121] Liao DZ, Pantazis CG, Hou X and Li SA. Promotion of estrogen-induced mammary gland carcinogenesis by androgen in the male Noble rat: probable mediation by steroid receptors. Carcinogenesis 1998; 19: 2173-2180.
- [122] Cochrane DR, Bernales S, Jacobsen BM, Cittelly DM, Howe EN, D'Amato NC, Spoelstra NS, Edgerton SM, Jean A, Guerrero J, Gomez F, Medicherla S, Alfaro IE, McCullagh E, Jedlicka P, Torkko KC, Thor AD, Elias AD, Protter AA and Richer JK. Role of the androgen receptor in breast cancer and preclinical analysis of enzalutamide. Breast Cancer Res 2014; 16: R7
- [123] McNamara KM, Moore NL, Hickey TE, Sasano H and Tilley WD. Complexities of androgen receptor signalling in breast cancer. Endocr Relat Cancer 2014; 21: T161-181.
- [124] Girault I, Andrieu C, Tozlu S, Spyratos F, Bieche I and Lidereau R. Altered expression pattern of alternatively spliced estrogen receptor beta transcripts in breast carcinoma. Cancer Lett 2004; 215: 101-112.
- [125] Shaaban AM, O'Neill PA, Davies MP, Sibson R, West CR, Smith PH and Foster CS. Declining estrogen receptor-beta expression defines ma-

- lignant progression of human breast neoplasia. Am J Surg Pathol 2003; 27: 1502-1512.
- [126] Mann S, Laucirica R, Carlson N, Younes PS, Ali N, Younes A, Li Y and Younes M. Estrogen receptor beta expression in invasive breast cancer. Hum Pathol 2001; 32: 113-118.
- [127] Murphy L, Cherlet T, Lewis A, Banu Y and Watson P. New insights into estrogen receptor function in human breast cancer. Ann Med 2003; 35: 614-631.
- [128] Honma N, Horii R, Iwase T, Saji S, Younes M, Takubo K, Matsuura M, Ito Y, Akiyama F and Sakamoto G. Clinical importance of estrogen receptor-beta evaluation in breast cancer patients treated with adjuvant tamoxifen therapy. J Clin Oncol 2008; 26: 3727-3734.
- [129] Nakopoulou L, Lazaris AC, Panayotopoulou EG, Giannopoulou I, Givalos N, Markaki S and Keramopoulos A. The favourable prognostic value of oestrogen receptor beta immunohistochemical expression in breast cancer. J Clin Pathol 2004; 57: 523-528.
- [130] Yan Y, Li X, Blanchard A, Bramwell VH, Pritchard KI, Tu D, Shepherd L, Myal Y, Penner C, Watson PH, Leygue E and Murphy LC. Expression of both estrogen receptor-beta 1 (ER-beta1) and its co-regulator steroid receptor RNA activator protein (SRAP) are predictive for benefit from tamoxifen therapy in patients with estrogen receptor-alpha (ER-alpha)-negative early breast cancer (EBC). Ann Oncol 2013; 24: 1986-1993.
- [131] Murphy LC and Leygue E. The role of estrogen receptor-beta in breast cancer. Semin Reprod Med 2012; 30: 5-13.
- [132] Chen Y, Chen L, Li JY, Mukaida N, Wang Q, Yang C, Yin WJ, Zeng XH, Jin W and Shao ZM. ERbeta and PEA3 co-activate IL-8 expression and promote the invasion of breast cancer cells. Cancer Biol Ther 2011; 11: 497-511.
- [133] Lin Y, Huang R, Chen L, Li S, Shi Q, Jordan C and Huang RP. Identification of interleukin-8 as estrogen receptor-regulated factor involved in breast cancer invasion and angiogenesis by protein arrays. Int J Cancer 2004; 109: 507-515.
- [134] Cappelletti V, Miodini P, Di Fronzo G and Daidone MG. Modulation of estrogen receptor-beta isoforms by phytoestrogens in breast cancer cells. Int J Oncol 2006; 28: 1185-1191.
- [135] Leygue E, Dotzlaw H, Watson PH and Murphy LC. Expression of estrogen receptor beta1, beta2, and beta5 messenger RNAs in human breast tissue. Cancer Res 1999; 59: 1175-1179.
- [136] Davies MP, O'Neill PA, Innes H, Sibson DR, Prime W, Holcombe C and Foster CS. Correlation of mRNA for oestrogen receptor beta

- splice variants ERbeta1, ERbeta2/ERbetacx and ERbeta5 with outcome in endocrine-treated breast cancer. J Mol Endocrinol 2004; 33: 773-782.
- [137] Shaaban AM, Green AR, Karthik S, Alizadeh Y, Hughes TA, Harkins L, Ellis IO, Robertson JF, Paish EC, Saunders PT, Groome NP and Speirs V. Nuclear and cytoplasmic expression of ERbeta1, ERbeta2, and ERbeta5 identifies distinct prognostic outcome for breast cancer patients. Clin Cancer Res 2008; 14: 5228-5235.
- [138] Chantzi NI, Tiniakos DG, Palaiologou M, Goutas N, Filippidis T, Vassilaros SD, Dhimolea E, Mitsiou DJ and Alexis MN. Estrogen receptor beta 2 is associated with poor prognosis in estrogen receptor alpha-negative breast carcinoma. J Cancer Res Clin Oncol 2013; 139: 1489-1498.
- [139] Leung YK, Lam HM, Wu S, Song D, Levin L, Cheng L, Wu CL and Ho SM. Estrogen receptor beta2 and beta5 are associated with poor prognosis in prostate cancer, and promote cancer cell migration and invasion. Endocr Relat Cancer 2010; 17: 675-689.
- [140] Esslimani-Sahla M, Simony-Lafontaine J, Kramar A, Lavaill R, Mollevi C, Warner M, Gustafsson JA and Rochefort H. Estrogen receptor beta (ER beta) level but not its ER beta cx variant helps to predict tamoxifen resistance in breast cancer. Clin Cancer Res 2004; 10: 5769-5776.
- [141] Hopp TA, Weiss HL, Parra IS, Cui Y, Osborne CK and Fuqua SA. Low levels of estrogen receptor beta protein predict resistance to tamoxifen therapy in breast cancer. Clin Cancer Res 2004; 10: 7490-7499.
- [142] Lee MT, Ho SM, Tarapore P, Chung I and Leung YK. Estrogen receptor beta isoform 5 confers sensitivity of breast cancer cell lines to chemotherapeutic agent-induced apoptosis through interaction with Bcl2L12. Neoplasia 2013; 15: 1262-1271.
- [143] Hamilton-Burke W, Coleman L, Cummings M, Green CA, Holliday DL, Horgan K, Maraqa L, Peter MB, Pollock S, Shaaban AM, Smith L and Speirs V. Phosphorylation of estrogen receptor beta at serine 105 is associated with good prognosis in breast cancer. Am J Pathol 2010; 177: 1079-1086.
- [144] Bernoux A, de Cremoux P, Lainé-Bidron C, Martin EC, Asselain B and Magdelénat H. Estrogen receptor negative and progesterone receptor positive primary breast cancer: pathological characteristics and clinical outcome. Institut curie breast cancer study group. Breast Cancer Res Treat 1998; 49: 219-225.
- [145] Nagai R, Kataoka M, Kobayashi S, Ishihara K, Tobioka N, Nakashima K, Naruse M, Saito K and Sakuma S. Estrogen and progesterone re-

- ceptors in human breast cancer with concomitant assay of plasma 17beta-estradiol, progesterone, and prolactin levels. Cancer Res 1979; 39: 1834-1840.
- [146] Rakha EA, El-Sayed ME, Green AR, Paish EC, Powe DG, Gee J, Nicholson RI, Lee AH, Robertson JF and Ellis IO. Biologic and clinical characteristics of breast cancer with single hormone receptor positive phenotype. J Clin Oncol 2007; 25: 4772-4778.
- [147] Allan GF and Sui Z. Non-steroidal progesterone receptor specific ligands. Mini Rev Med Chem 2005; 5: 701-708.
- [148] Colditz GA, Rosner BA, Chen WY, Holmes MD and Hankinson SE. Risk factors for breast cancer according to estrogen and progesterone receptor status. J Natl Cancer Inst 2004; 96: 218-228.
- [149] Coombes RC, Hall E, Gibson LJ, Paridaens R, Jassem J, Delozier T, Jones SE, Alvarez I, Bertelli G, Ortmann O, Coates AS, Bajetta E, Dodwell D, Coleman RE, Fallowfield LJ, Mickiewicz E, Andersen J, Lønning PE, Cocconi G, Stewart A, Stuart N, Snowdon CF, Carpentieri M, Massimini G, Bliss JM, van de Velde C; Intergroup Exemestane Study. A randomized trial of exemestane after two to three years of tamoxifen therapy in postmenopausal women with primary breast cancer. N Engl J Med 2004; 350: 1081-1092.
- [150] Dowsett M, Dixon JM, Horgan K, Salter J, Hills M and Harvey E. Antiproliferative effects of idoxifene in a placebo-controlled trial in primary human breast cancer. Clin Cancer Res 2000; 6: 2260-2267.
- [151] Johnston SR, Saccani-Jotti G, Smith IE, Salter J, Newby J, Coppen M, Ebbs SR and Dowsett M. Changes in estrogen receptor, progesterone receptor, and pS2 expression in tamoxifen-resistant human breast cancer. Cancer Res 1995; 55: 3331-3338.
- [152] Mason BH, Holdaway IM, Mullins PR, Yee LH and Kay RG. Progesterone and estrogen receptors as prognostic variables in breast cancer. Cancer Res 1983; 43: 2985-2990.
- [153] De Maeyer L, Van Limbergen E, De Nys K, Moerman P, Pochet N, Hendrickx W, Wildiers H, Paridaens R, Smeets A, Christiaens MR, Vergote I, Leunen K, Amant F and Neven P. Does estrogen receptor negative/progesterone receptor positive breast carcinoma exist? J Clin Oncol 2008; 26: 335-336; author reply 336-338.
- [154] Hefti MM, Hu R, Knoblauch NW, Collins LC, Haibe-Kains B, Tamimi RM and Beck AH. Estrogen receptor negative/progesterone receptor positive breast cancer is not a reproducible subtype. Breast Cancer Res 2013; 15: R68.

- [155] Nadji M, Gomez-Fernandez C, Ganjei-Azar P and Morales AR. Immunohistochemistry of estrogen and progesterone receptors reconsidered: experience with 5,993 breast cancers. Am J Clin Pathol 2005; 123: 21-27.
- [156] Iwase H, Greenman JM, Barnes DM, Hodgson S, Bobrow L and Mathew CG. Sequence variants of the estrogen receptor (ER) gene found in breast cancer patients with ER negative and progesterone receptor positive tumors. Cancer Lett 1996; 108: 179-184.
- [157] Auchus RJ and Fuqua SA. Prognostic factors and variant estrogen receptor RNAs in clinical breast cancer. Nucl Med Biol 1994; 21: 449-454.
- [158] Daffada AA, Johnston SR, Smith IE, Detre S, King N and Dowsett M. Exon 5 deletion variant estrogen receptor messenger RNA expression in relation to tamoxifen resistance and progesterone receptor/pS2 status in human breast cancer. Cancer Res 1995; 55: 288-293.
- [159] Fuqua SA, Allred DC and Auchus RJ. Expression of estrogen receptor variants. J Cell Biochem Suppl 1993; 17G: 194-197.
- [160] Schultz JR, Petz LN and Nardulli AM. Estrogen receptor alpha and Sp1 regulate progesterone receptor gene expression. Mol Cell Endocrinol 2003; 201: 165-175.
- [161] Rickard DJ, Waters KM, Ruesink TJ, Khosla S, Katzenellenbogen JA, Katzenellenbogen BS, Riggs BL and Spelsberg TC. Estrogen receptor isoform-specific induction of progesterone receptors in human osteoblasts. J Bone Miner Res 2002; 17: 580-592.
- [162] Shen Q, Zhang Y, Uray IP, Hill JL, Kim HT, Lu C, Young MR, Gunther EJ, Hilsenbeck SG, Chodosh LA, Colburn NH and Brown PH. The AP-1 transcription factor regulates postnatal mammary gland development. Dev Biol 2006; 295: 589-603.
- [163] Encarnación CA, Ciocca DR, McGuire WL, Clark GM, Fuqua SA and Osborne CK. Measurement of steroid hormone receptors in breast cancer patients on tamoxifen. Breast Cancer Res Treat 1993; 26: 237-246.
- [164] Colomer R, Beltran M, Dorcas J, Cortes-Funes H, Hornedo J, Valentin V, Vargas C, Mendiola C and Ciruelos E. It is not time to stop progesterone receptor testing in breast cancer. J Clin Oncol 2005; 23: 3868-3869; author reply 3869-3870.
- [165] Keshgegian AA and Cnaan A. Estrogen receptor-negative, progesterone receptor-positive breast carcinoma: poor clinical outcome. Arch Pathol Lab Med 1996; 120: 970-973.
- [166] Ng CH, Pathy NB, Taib NA, Mun KS, Rhodes A and Yip CH. The estrogen receptor negativeprogesterone receptor positive breast carcinoma is a biological entity and not a technical

- artifact. Asian Pac J Cancer Prev 2012; 13: 1111-1113.
- [167] Bardon S, Vignon F, Montcourrier P and Rochefort H. Steroid receptor-mediated cytotoxicity of an antiestrogen and an antiprogestin in breast cancer cells. Cancer Res 1987; 47: 1441-1448.
- [168] Bardon S, Vignon F, Chalbos D and Rochefort H. RU486, a progestin and glucocorticoid antagonist, inhibits the growth of breast cancer cells via the progesterone receptor. J Clin Endocrinol Metab 1985; 60: 692-697.
- [169] Bray JD, Zhang Z, Winneker RC and Lyttle CR. Regulation of gene expression by PRA-910, a novel progesterone receptor modulator, in T47D cells. Steroids 2003; 68: 995-1003.
- [170] Bowden RT, Hissom JR and Moore MR. Growth stimulation of T47D human breast cancer cells by the anti-progestin RU486. Endocrinology 1989; 124: 2642-2644.
- [171] Hissom JR and Moore MR. Progestin effects on growth in the human breast cancer cell line T-47D-possible therapeutic implications. Biochem Biophys Res Commun 1987; 145: 706-711.
- [172] Jeng MH, Langan-Fahey SM and Jordan VC. Estrogenic actions of RU486 in hormone-responsive MCF-7 human breast cancer cells. Endocrinology 1993; 132: 2622-2630.
- [173] Jonat W, Bachelot T, Ruhstaller T, Kuss I, Reimann U and Robertson JFR. Randomized phase II study of lonaprisan as second-line therapy for progesterone receptor-positive breast cancer. Ann Oncol 2013; 24: 2543-2548.
- [174] Klijn JG, de Jong FH, Bakker GH, Lamberts SW, Rodenburg CJ and Alexieva-Figusch J. Antiprogestins, a new form of endocrine therapy for human breast cancer. Cancer Res 1989; 49: 2851-2856.
- [175] Perrault D, Eisenhauer EA, Pritchard KI, Panasci L, Norris B, Vandenberg T and Fisher B. Phase II study of the progesterone antagonist mifepristone in patients with untreated metastatic breast carcinoma: a National Cancer Institute of Canada Clinical Trials Group study. J Clin Oncol 1996: 14: 2709-2712.
- [176] El Etreby MF, Liang Y, Wrenn RW and Schoenlein PV. Additive effect of mifepristone and tamoxifen on apoptotic pathways in MCF-7 human breast cancer cells. Breast Cancer Res Treat 1998; 51: 149-168.
- [177] Lee JY, Shin JY, Kim HS, Heo JI, Kho YJ, Kang HJ, Park SH and Lee JY. Effect of combined treatment with progesterone and tamoxifen on the growth and apoptosis of human ovarian cancer cells. Oncol Rep 2012; 27: 87-93.
- [178] Nishino T, Ishibashi K, Hirtreiter C and Nishino Y. Potentiation of the antitumor effect of

Steroid receptors and breast cancer

- tamoxifen by combination with the antiprogestin onapristone. J Steroid Biochem Mol Biol 2009; 116: 187-190.
- [179] Agoff SN, Swanson PE, Linden H, Hawes SE and Lawton TJ. Androgen receptor expression in estrogen receptor-negative breast cancer. Immunohistochemical, clinical, and prognostic associations. Am J Clin Pathol 2003; 120: 725-731.
- [180] Park S, Koo J, Park HS, Kim JH, Choi SY, Lee JH, Park BW and Lee KS. Expression of androgen receptors in primary breast cancer. Ann Oncol 2010; 21: 488-492.
- [181] Doane AS, Danso M, Lal P, Donaton M, Zhang L, Hudis C and Gerald WL. An estrogen receptor-negative breast cancer subset characterized by a hormonally regulated transcriptional program and response to androgen. Oncogene 2006; 25: 3994-4008.
- [182] Ni M, Chen Y, Lim E, Wimberly H, Bailey ST, Imai Y, Rimm DL, Liu XS and Brown M. Targeting androgen receptor in estrogen receptornegative breast cancer. Cancer Cell 2011; 20: 119-131.
- [183] He J, Peng R, Yuan Z, Wang S, Peng J, Lin G, Jiang X and Qin T. Prognostic value of androgen receptor expression in operable triple-negative breast cancer: a retrospective analysis based on a tissue microarray. Med Oncol 2012; 29: 406-410.
- [184] Rakha EA, El-Sayed ME, Green AR, Lee AH, Robertson JF and Ellis IO. Prognostic markers in triple-negative breast cancer. Cancer 2007; 109: 25-32.
- [185] Lehmann BD, Bauer JA, Chen X, Sanders ME, Chakravarthy AB, Shyr Y and Pietenpol JA. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. J Clin Invest 2011; 121: 2750-2767.
- [186] Tang D, Xu S, Zhang Q and Zhao W. The expression and clinical significance of the androgen receptor and E-cadherin in triple-negative breast cancer. Med Oncol 2012; 29: 526-533.
- [187] Hardin C, Pommier R, Calhoun K, Muller P, Jackson T and Pommier S. A new hormonal therapy for estrogen receptor-negative breast cancer. World J Surg 2007; 31: 1041-1046.

- [188] Arce-Salinas C, Riesco-Martinez MC, Hanna W, Bedard P and Warner E. Complete response of metastatic androgen receptor-positive breast cancer to bicalutamide: case report and review of the literature. J Clin Oncol 2016; 34: e21-4.
- [189] Desmedt C, Voet T, Sotiriou C and Campbell PJ. Next-generation sequencing in breast cancer: first take home messages. Curr Opin Oncol 2012; 24: 597-604.
- [190] Elshimali YI, Khaddour H, Sarkissyan M, Wu Y and Vadgama JV. The clinical utilization of circulating cell free DNA (CCFDNA) in blood of cancer patients. Int J Mol Sci 2013; 14: 18925-18958.
- [191] Mirza S, Sharma G, Parshad R, Srivastava A, Gupta SD and Ralhan R. Clinical significance of promoter hypermethylation of ERbeta and RARbeta2 in tumor and serum DNA in Indian breast cancer patients. Ann Surg Oncol 2012; 19: 3107-3115.
- [192] Russnes HG, Navin N, Hicks J and Borresen-Dale AL. Insight into the heterogeneity of breast cancer through next-generation sequencing. J Clin Invest 2011; 121: 3810-3818.
- [193] Tung N, Battelli C, Allen B, Kaldate R, Bhatnagar S, Bowles K, Timms K, Garber JE, Herold C, Ellisen L, Krejdovsky J, DeLeonardis K, Sedgwick K, Soltis K, Roa B, Wenstrup RJ and Hartman AR. Frequency of mutations in individuals with breast cancer referred for BRCA1 and BRCA2 testing using next-generation sequencing with a 25-gene panel. Cancer 2015; 121: 25-33.
- [194] Levenson RM, Borowsky AD and Angelo M. Immunohistochemistry and mass spectrometry for highly multiplexed cellular molecular imaging. Lab Invest 2015; 95: 397-405.
- [195] Angelo M, Bendall SC, Finck R, Hale MB, Hitzman C, Borowsky AD, Levenson RM, Lowe JB, Liu SD, Zhao S, Natkunam Y and Nolan GP. Multiplexed ion beam imaging of human breast tumors. Nat Med 2014; 20: 436-442.
- [196] Trevino LS and Weigel NL. Phosphorylation: a fundamental regulator of steroid receptor action. Trends Endocrinol Metab 2013; 24: 515-524.